THE NEURAMINIDASE OF MEASLES VIRUS

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The agent of measles (rubeola) has been assumed to be a paramyxovirus on the basis of its electron microscopic appearance, its size, its sensitivity to ether and detergents, and the presence of hemagglutinin in the viral envelope. Thus far, however, the one attribute common to myxoviruses, namely neuraminidase, has not been described for measles virus. We have demonstrated that specific neuraminidase activity is associated with measles viral hemagglutinin. The evidence suggests strongly that the enzyme is part of the viral envelope.

MATERIALS AND METHODS

Measles virus (Edmonston strain) was propagated in HeLa or amnion cell monolayers grown in 32 oz prescription bottles. After adsorption of appropriately diluted virus onto washed monolayers for 1 hr at 37°C, cells were maintained for 6 to 7 days in Eagle's basal medium supplemented with 0.1% lactalbumin hydrolysate (LH) without serum. Cells were then scraped into the fluid and concentrated by centrifugation at 1500 X g; the pellet was frozen and thawed successively three times in a small volume of culture fluid, which, after

low speed clarification, was added to the remaining volume of culture fluid. The whole was then centrifuged at 33,000 X g for 2 hr at 4°C in a Spinco model L ultracentrifuge. The pelleted virus was taken up in phosphate-buffered saline, pH 7.2, to effect a 10- to 20-fold concentration of hemagglutinin (HA) and infectivity.

Hemagglutination-inhibition tests were performed by the pattern technique with 2% rhesus cells, using four hemagglutinating units (HAU) of virus. Infectivity titrations were performed by inoculating decimal dilutions of virus onto replicate amnion cell monolayers. After 3 days' incubation at 37°C under serum-free medium, cells were examined for hemadsorption of rhesus erythrocytes (0.5%). Neuraminidase activity was determined in 0.1 ml of concentrated virus added to an equal volume of substrate [(Collocalia mucoid (Howe et al. 1961), neuraminlactose (NAL) (Calbiochem), or urinary mucoprotein (UMP)] in pH 5.5 phosphate buffer. After incubation at 37°C for 16 to 18 hr, free N-acetylneuraminic acid (NANA) was measured by the thiobarbituric acid method (Warren, 1959). Controls (virus alone, substrate alone, and homogenates of normal cells) gave negligible color values. NANA was identified by descending chromatography of the reaction mixtures on Whatman no. 1 paper using isobutyric acidwater (5:3), adjusted to pH 3.7 with concentrated NH,OH, as solvent (Magasanik et al. 1950). Spots were developed by the thiobarbituric acid reaction (Warren, 1960), with crystalline NANA as reference standard. Absorption curves were plotted with a Cary model 11M recording spectrophotometer.

RESULTS AND DISCUSSION

Incubation of concentrated measles virus (64-640 HAU)

TABLE I

		μg		
	HAU	Substrateb		
Cell	virus	Collocalia	NAL	nM NANA
line	added ^a	mucoid		liberated ^C
HeLa	640	500		29
Amnion	320	200		17
			25	23
Amnion	640	200		28 ^d
			100	75 ^đ
HeLa	64	<u>UMP</u> e 202		17
	line HeLa Amnion Amnion	Cell virus line added ^a HeLa 640 Amnion 320 Amnion 640	HAU Substrate ^b Cell virus Collocalia line added ^a mucoid HeLa 640 500 Amnion 320 200 Amnion 640 200 UMPe	HAU Substrate b Cell virus Collocalia NAL line added mucoid HeLa 640 500 Amnion 320 200 25 Amnion 640 200 UMPe

Gave negligible color value in absence of substrate.

with either Collocalia mucoid or neuraminlactose at pH 5.5 at 37°C resulted in the liberation of thiobarbituric acid chromophore (Table I). Increasing the amount of substrate with a constant amount of virus resulted in correspondingly increased chromophore formation. The chromophore was identified as NANA by its characteristic absorption peak at 549 mu and by descending paper chromatography, which showed its

b Gave negligible color value in absence of virus.

C 37°C for 18 hr.

Descending chromatogram gave single spot with Rf of crystalline NANA.

e Urinary Mucoprotein: 110µg inhibited 4 HAU.

mobility to be the same as that of crystalline NANA. Urinary mucoprotein (UMP), a potent inhibitor of myxovirus hemagglutinin, was also susceptible to the viral enzyme. neuraminidases of other paramyxoviruses (Darrell and Howe, 1964; Tosawa et al. 1967) appear to be strain specific. Serologic analysis of measles virus neuraminidase is in progress. The foregoing results constitute the first reported demonstration that measles virus possesses a neuraminidase and further justify its designation as a paramyxovirus.

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